Synergistic action of bacteriophage with antibiotics on Pseudomonas aeruginosa biofilms

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ABSTRACT. An opportunistic microorganism Pseudomonas aeruginosa causes nosocomial infections and often forms rigid biofilms on both intact human skin and wounds, as well as the surfaces of artificial implants, prostheses and catheters. Being embedded into the exopolymERIC matrix of the biofilm bacteria are extremely resistant to antibiotics, biocides, and the immune system of the human. Therefore, the search for new approaches for eradication of bacterial biofilm is an urgent challenge of modern medicine. Combined therapy of biofilms with antibiotics and phages has shown promising results. In this study, 48-hour monomicrobial biofilms of Pseudomonas aeruginosa were treated with phage from the family Myoviridae isolated from Lake Baikal and 12 antibiotics. While the treatment with a single phage did not significantly affect neither planktonic nor biofilm-embedded bacteria, in the presence of the phage, the MICs of gentamicin, ciprofloxacin, imipenem and meropenem the biofilm decreased 4-8-fold. Moreover, the density of biofilm was also significantly dropped in compare with the treatment by solely antibiotics or phage.

Keywords: Pseudomonas aeruginosa, microbial biofilms, bacteriophages, antibiotics, synergism

1. Introduction

Biofilm formation is believed as one of the main factors leading to chronic P. aeruginosa infections (Bjarnsholt, 2013). Biofilms are communities of microbial cells immersed into the polymeric matrix formed on the surface of tissues, medical devices, catheters, and plastic implants. The bacteria in the biofilm are highly resistant to biocides, antibiotics and the human immune system, causing chronic reinfections and inflammatory complications in the postoperative period (Hilbert and Moore, 2005).

Bacteriophages (or phages), viruses infecting the bacteria, are natural antibacterial agents that specifically target and lyse bacteria. Some phages can penetrate the inner layers of biofilms and infect dormant cells (Pires et al., 2017), which is a clear advantage of phages over antibiotics in treatment of biofilms. Some studies have shown the efficiency of antibiotics combinations with phages on liquid cultures and biofilms of P. aeruginosa (Chaudhry et al., 2017). The advantage of this approach is 1) the low possibility of simultaneous development of resistance to both agents and 2) the effective concentrations of antibacterial agents are decreased several fold providing lowered risks of side effects.

2. Materials and methods

Pseudomonas aeruginosa ATCC ® 27853 ™ and the bacteriophage belonging to Myoviridae family, isolated from Lake Baikal and termed PaSA1 were used in this study. To analyze the biofilm, the bacteria were grown under static conditions at 35 °C for 48 hours in BM broth (Kayumov et al., 2015; Baidamshina et al., 2017). Biofilm formation was evaluated in 24-well polystyrene plates (Eppendorf) by staining with crystal violet (Sharafutdinov et al., 2017). The minimal inhibitory concentration (MIC) of antimicrobials was determined by broth microdilution in accordance with the EUCAST recommendation. Cell viability was assessed using confocal laser scanning microscopy and a resasurine assay.

3. Results and discussion

Among various bacteriophages from Lake Baikal in the area of wastewaters shedding close to Slyudyanka city, the isolate PaSA1 was capable of lysing P. aeruginosa ATCC 27853 cells. By using the transmission electron microscope the phage has been identified according to morphology as a member of the Myoviridae family. Next, the synergistic effect of the bacteriophage with 12 different antibiotics was studied: gentamicin, amikacin,
ciprofloxacin, norfloxacin, ceftriaxone, streptomycin, erythromycin, azithromycin, vancomycin, imipenem, meropenem and colistin. In combination with phage, the MIC of gentamicin, ciprofloxacin, imipenem and meropenem were 4-8 times lower in compare with solely antibiotic. The synergistic effect of these drugs and the phage was also observed on 48-h old biofilms. Taken together, our data allow suggesting PaSA1 as promising tool for enhancement of antimicrobial efficacy against P. aeruginosa including its biofilms.

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