Representatives of Diplomonadida in fishes of East Siberia

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ABSTRACT. In this study, we analyze the diversity of flagellated protozoa Diplomonadida, which are parasites in the digestive system of animals, in fishes of East Siberia. We have developed a method to diagnose the presence of DNA of Diplomonadida representatives, in particular, of Spironucleus barkhanus, in the analyzed samples. In our previous studies, we used molecular genetic methods to analyze salmonids of East Siberia for the presence of diplomonads and to determine the infection. The analysis of genetic diversity of diplomonads in the fishes of the genera Coregonus and Thymallus revealed the presence of one species, S. barkhanus. However, in addition to a cosmopolitan genotype widespread among Holarctic salmonids, in C. migratorius and C. lavaretus baikalensis, we have found a new genotype obviously different from those registered previously. Probably, S. barkhanus of this genotype is endemic for Lake Baikal. To determine a probable parasite exchange between C. migratorius and other fish species in the pelagic zone of Lake Baikal, we performed a screening of near-shore pelagic and pelagic cottoid fish for the presence of S. barkhanus. The analysis did not indicate the presence of DNA of S. barkhanus in the studied samples. We suppose the presence of other diplomonads species in endemic cottoid fish different from S. barkhanus.

Keywords: molecular genetic methods, Lake Baikal, Coregonus, Thymallus, Comephorus, Cottocomephorus

Diplomonadida are the flagellated protozoa that for many years have been considered the most primitive of eukaryotes, since they lack the typical mitochondria, peroxisomes and the Golgi apparatus (Keeling and Doolittle, 1997). The discovery of several varieties of mitochondria in Diplomonadida revealed that their cells were secondarily simplified, and these protists cannot be an intermediate stage in the formation of organelles in the general scheme of the evolutionary history of eukaryotes (Jerlstrom-Hultqvist et al., 2013). The order Diplomonadida includes the genera Spironucleus, Hexamita, Treponomonas, Trimitus, Enteromonas, Octomitus, and Giardia (Kolisko et al., 2008). The species Hexamita are mostly free-living anaerobic organisms inhabiting bottom sediments. At the same time, other taxa are almost purely commensals and pathogens, which usually inhabit the digestive system of mammals, birds, reptiles, amphibians, arthropods, molluscs, and fishes (Williams et al., 2011).

Diplomonads are of special interest for veterinary and agricultural organisations due to the severe pathologies that they cause in bred animals. Species of the genus Spironucleus are of particular attention, since they can cause devastating episodes of systemic infections in both ornamental and commercial fishes. These ubiquitous flagellates are found in cold, temperate and tropical waters and can infect a wide range of freshwater and marine fishes, as well as molluscs and crustaceans (Noga, 2010). The fish industry (and aquaculture in general) is becoming increasingly important in the world, since the natural fish stocks are depleted. At the same time, fishes and other objects of aquaculture are especially susceptible to outbreaks of diseases caused by diplomonads. For instance, Spironucleus salmonicida Jorgensen & Sterud, 2006 caused massive episodes of systemic infections in the farm-raised Salmo salar Linnaeus, 1758 and Oncorhynchus tschawytscha (Walbaum, 1792) (Kent et al., 1992; Poppe and Mo, 1993). The life cycle of diplomonads is direct and includes the stage of mobile parasitic trophozoite, which is generally followed by a stage of cysts. Infection rate is extremely high, especially in aquaculture, which also leads to a significantly high risk of host death.

The biology and ecology of species of the genus Spironucleus are poorly investigated in many aspects, including specific hosts, geographic ranges, transmission mechanisms, and pathogenicity of different species. Lack of knowledge is a serious limitation in disease control, since it impedes an accurate diagnosis and identification of the infection source. These obstacles are primarily associated with difficult detection of the genera and species of the order Diplomonadida,

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which also explains the confusion in the nomenclature. The descriptions of the diplomonads are incomplete. Morphological studies using scanning and transmission microscopy often do not allow accurate identification of species. For example, *Spironucleus barkhanus* Sterud, Mo, & Poppe, 1998 has been described for a long time as eukaryotic organism of two morphologically similar types (nonpathogenic freshwater and pathogenic marine) (Sterud et al., 1998). Based only on the molecular genetic analysis, the representatives of this pathogenic genotype were described as a new species (*S. salmonicida*). Subsequently, the gene sequences of the small subunit ribosomal DNA were determined for all known species of diplomonads isolated from fishes and belonging to the genus *Spironucleus*: *S. barkhanus*, *S. salmonicida*, *S. baicalensis*, and *S. vortens*. Previously, they were studied by ultrastructure analysis (Jørgensen and Sterud, 2006). Therefore, molecular genetic analysis plays a key role in detection and identification of diplomonads in fishes.

Despite the relatively high level of knowledge about animals of Lake Baikal and water bodies of East Siberia, data on the species composition of parasitic protozoa are still fragmentary and incomplete. Previously, representatives of the genus *Hexamita* were described by morphological characteristics in *Coregonus migratorius* (Georgi, 1775), *Thymallus baicalensis* Dybowski, 1874, *Thymallus brevippinis* Svetovidov, 1931, *Leuciscus baikalensis* (Dybowski, 1874), *Lota lota* (Linnaeus, 1758), and endemic cottoid fish: *Batrachochottus multiradiatus* Berg, 1907, *Batrachocottus nikolskii* (Berg, 1900), *Cottocomephorus grewingkii* (Dybowski, 1874), and *Limnocottus bergianus* (Taliev, 1935) (Zaika, 1965; Pronin, 2001; Pugachev, 2001; Rusinek, 2007).

Nucleotide sequences of a gene fragment of the small subunit ribosomal RNA were obtained in the study of the microbeita from the intestinal tract of *T. baikalensis* from the Angara River. These sequences were identical to *S. barkhanus* from *Thymallus thymallus* (Linnaeus, 1758) and *Salvelinus alpinus* (Linnaeus, 1758) (Belkova et al., 2008; 2009). Based on the sequences obtained, a diagnostic system was developed allowing detection of diplomonads and determination of their species (genotype).

In this study, we performed molecular genetic studies by standard methods: DNA was extracted using a DNA-Sorb-B kit according to the manufacturer’s instructions (Ampli-Sens, Moscow); PCR was carried out in a standard reaction mixture.

The regions conservative for eukaryotes (DpFun-DpR) served as primer to diagnose the presence of DNA of diplomonads. At the same time, the difference in the length of the observed gene fragment in diplomonads (~450 bp) and other eukaryotic organisms (~630 and more) provided the specificity of the target amplicons. Gene fragments of the small subunit rRNA of *S. barkhanus* were amplified using species-specific primers developed and tested in this study (Table 1, Fig. 1).

Using the developed diagnostic system, we determined infection in the representatives of the genera *Coregonus* and *Thymallus* (Denikina et al., 2011; 2016; 2017). The representatives of the genus *Thymallus* occupy a large area of unconnected basins of the Palaeartic and Nonarctic rivers and lakes. Analysis of fish of the genus *Thymallus* from water bodies of East Siberia and Mongolia has shown an obvious trend in infection of graylings *S. barkhanus* in the catchment area of the Angara River: the minimum in Lake Khubsugul (38.5 %), the significant amount in the Barguzin River and Lake Baikal (80.0 and 85.2 %, respectively) and the maximum in the Angara River (100 %) (Denikina et al., 2011).

The genus *Coregonus* in water bodies of southeastern Siberia is represented by species that occupy different ecological niches and have different food strategies. *S. barkhanus* infection rate in *C. migratorius* varies from 29.3 to 76.2 %. *Coregonus lavaretus baikalensis* Dybowski, 1874 is slightly infected (10-20%). The infection rate of *Coregonus* species in the Barguzin River was 12.5 %, in the tributaries of the Lena River is even lower. Therefore, habitat conditions, food preferences, behaviour and morphological features of the jaw structure are the determining factors of infection (Denikina et al., 2016). Analysis of the genetic diversity of diplomonads in fish of the genera *Coregonus* and *Thymallus* revealed

![Table 1. Primer pairs used in the study](image)

<table>
<thead>
<tr>
<th>Direct primer sequence</th>
<th>Reverse primer sequence</th>
<th>Amplicon length (bp)</th>
</tr>
</thead>
<tbody>
<tr>
<td>DPFUN 5’-GCCAGCAGCGCGGTAAATTCC</td>
<td>DPR 5’-AGCCGCAGACTCCACRTCT</td>
<td>450</td>
</tr>
<tr>
<td>DPSF 5’- CAGCCCGGGTTAAATTCGACAC</td>
<td>DPR1+ 5’-AGCCGCAGCTCCATCGTGTTG</td>
<td>450</td>
</tr>
<tr>
<td>DPSF 5’- CAGCCCGGGTTAAATTCGACAC</td>
<td>Sp+R 5’-GCAGCCTTGTACGACTTCC</td>
<td>984</td>
</tr>
<tr>
<td>Sp+1F 5’-GCCATGCATGCTATGTGTAGAC</td>
<td>DpR1+ 5’-AGCCGCAGACTCCAGTCTGTTG</td>
<td>881</td>
</tr>
</tbody>
</table>
the presence of a single species of *S. barkhanus*. At the same time, along with the cosmopolitan genotype, which is widespread in salmonid fish of Holarctic, in *C. migratorius* and *C. lavaretus baicalensis* we found a new genotype significantly different from all previously recorded ones. *S. barkhanus* of this genotype is likely to be endemic for Lake Baikal (Denikina et al., 2016; 2017).

To identify a possible parasite exchange between *C. migratorius* and other fish species in the pelagic zone of Lake Baikal, we screened the coastal-pelagic and pelagic cottoid fish for the presence of *S. barkhanus*. To investigate the infestation of endemic species of cottoid fish, we collected the material from 25 May to 15 June 2011 throughout the entire water area of Lake Baikal. The fishes were collected with an RK-15/30 midwater trawl at different depths. In total, we investigated 27 specimens of *Comephorus dybowski* Korotneff, 1905, 5 specimens of *Comephorus baicalensis* (Pallas, 1776), 11 specimens of *Cottocomephorus inermis* (Jakowlew, 1890), 58 specimens of *Cottocomephorus grewingkii* (Dybowskii, 1874), and 60 specimens of *Cottocomephorus alexandrae* Taliev, 1935 (Table 2, Fig. 2).

The necessity of these experiments was due to the following circumstances. i) Along with *C. migratorius*, five endemic Cottoidei species inhabit the open pelagic zone of Lake Baikal: pelagic species (*C. baicalensis* and *C. dybowski*) and three bentheopelagic species of the genus *Cottocomephorus* (*C. inermis*, *C. grewingkii* and *C. alexandrae*). The habitats of these species coincide with those of *C. migratorius* in the winter and spring period. ii) The food spectra of these species are similar with the food spectrum of *C. migratorius* in the feeding period and includes zooplankton: *Epischura baikalensis* Sars, 1900 and *Macrohectopus branickii* (Dybowsky, 1874). iii) *T. baicalensis* inhabit the coastal zone of Lake Baikal, where bentopelagic cottoid fish spawn. There is evidence of the finding of *Hexamita* in *C. grewingkii* (Zaika, 1965). Since species determined in 1965 is rather ambiguous considering the revised classification, and the indicated localization (the intestine and gallbladder) corresponds to *S. barkhanus*, the analysis was necessary. All above-mentioned ecological features of cottoid fish inhabiting the pelagic zone of the lake suggest that they have diplomonads, which were previously found in *T. baicalensis* and *C. migratorius*. However, the analysis did not reveal the presence of *S. barkhanus* in the studied DNA samples. The presence of *Hexamita* in *C. grewingkii* (Zaika, 1965) and the absence of *S. barkhanus* in our samples can be explained by the fact that the fish studied differed greatly. Most likely, in (Zaika, 1965) fish for parasitological analysis were collected from the coastal zone of Lake Baikal. *C. grewingkii* is the most numerous representative of the shallow water ichthyocenosis. Within a species, three spawning populations (schools) are distinguished at different times: March, May, and August (Taliev, 1955; Koryakov, 1972). Multi-layer rocky grounds in the coastal zone of the lake serve for *C. grewingkii* as a substrate for spawning. In schools of *C. grewingkii* spawning at different times, the incubation period for the development of clutches

![Fig. 2. Schematic sample collection map, May-June 2011](image)

<table>
<thead>
<tr>
<th>Trawl coordinates</th>
<th>Fish species</th>
<th>Quantity, specimen</th>
</tr>
</thead>
<tbody>
<tr>
<td>51°51’17”N 104°37’39”E</td>
<td><em>C. dybowski</em></td>
<td>5</td>
</tr>
<tr>
<td>52°03’67”N 106°02’15”E</td>
<td><em>C. baicalensis</em></td>
<td>3</td>
</tr>
<tr>
<td>52°08’94”N 108°81’32”E</td>
<td><em>C. dybowski</em></td>
<td>5</td>
</tr>
<tr>
<td>52°25’46”N 106°02’41”E</td>
<td><em>C. grewingkii</em></td>
<td>10</td>
</tr>
<tr>
<td>52°46’53”N 106°67’59”E</td>
<td><em>C. inermis</em></td>
<td>3</td>
</tr>
<tr>
<td>53°43’39”N 108°68’59”E</td>
<td><em>C. inermis</em></td>
<td>3</td>
</tr>
<tr>
<td>53°46’57”N 107°60’67”E</td>
<td><em>C. inermis</em></td>
<td>3</td>
</tr>
<tr>
<td>52°38’64”N 106°18’06”E</td>
<td><em>C. alexandrae</em></td>
<td>30</td>
</tr>
<tr>
<td>52°31’85”N 106°05’97”E</td>
<td><em>C. grewingkii</em></td>
<td>20</td>
</tr>
<tr>
<td></td>
<td><em>C. inermis</em></td>
<td>1</td>
</tr>
<tr>
<td></td>
<td><em>C. alexandrae</em></td>
<td>20</td>
</tr>
<tr>
<td></td>
<td><em>C. dybowski</em></td>
<td>12</td>
</tr>
</tbody>
</table>

Table 2. Specimens of cottoid fish, Lake Baikal May-June 2011
ranges from 20 to 80-90 days depending on spawning season and temperature conditions during the development of eggs. Male protects the clutch from the moment of spawning to the appearance of larvae. Most males die after the period of clutch protection. This zone is the main feeding ground for the *T. baikalensis*, which infection rate reaches 85.2 %. We assumed that *C. grewingkii* samples with *Hexamita* (Zaika, 1965) were collected in the spawning area. Therefore, we did not detect *S. barkhanus* in the open pelagic zone of the lake in the fattening *C. grewingkii* specimens. It is also likely that diplomonads in salmonid and cottoid fish were attributed to the same species by mistake, since previous data by (Zaika, 1965) were obtained using methods of light microscopy. In this case, cottoid fish may have other diplomonads species than *S. barkhanus*. This assumption requires additional studies.

**Acknowledgments**

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**References**


